



## Highly enantioselective fluorescent sensor for chiral recognition of amino acid derivatives

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### ABSTRACT

A highly enantioselective fluorescent sensor, containing benzylaminomethyl groups at 3,3'-position of 1,1'-bi-2-naphthol (BINOL), has been used to conduct the chiral recognition of  $\alpha$ -amino acid derivatives. It is observed that one enantiomer of *N*-Boc-proline can increase the fluorescence intensity of the binaphthyl fluorophores by over 57-fold, while the other enantiomer can cause only sixfold fluorescence enhancement. Such unusually highly enantioselective response demonstrates that this sensor is potentially useful in the enantioselective recognition of amino acid derivatives.

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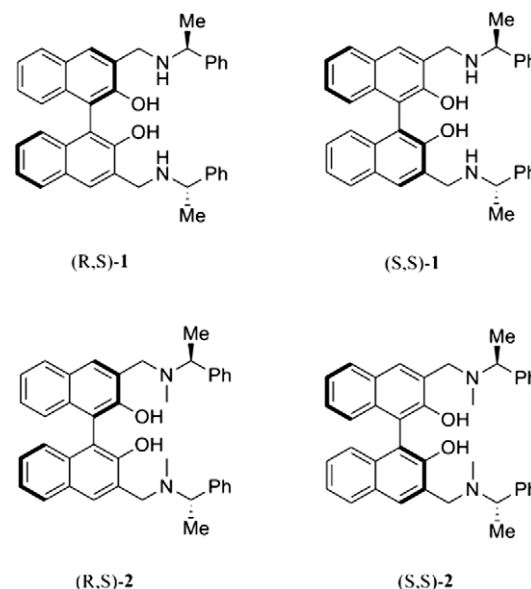
Molecular recognition, especially chiral recognition, is one of the most fundamental and crucial properties of various natural systems.<sup>1</sup> It is of particular significance for understanding the interactions of biological molecules and for the designing of asymmetric catalysis systems.<sup>2</sup> With the increasingly successful application to smart artificial systems, chiral recognition which is a broader and more commonly used 'tool' for academic research and industry<sup>3</sup> has gained greater scientific maturity.

Amino acids and their derivatives play an important role in a wide variety of biological processes.<sup>4</sup> A number of studies based on various analytical methods for chiral amino acids recognition have been reported, such as capillary electrophoresis,<sup>5</sup> HPLC,<sup>6</sup> absorption spectrometry,<sup>7</sup> and electrochemistry.<sup>8</sup> Fluorescence spectroscopy has only limited reports on chiral amino acid recognition.<sup>9</sup> In 2000, Corradini and co-workers prepared the  $\alpha$ -cyclodextrin-based sensor molecules, which could be enantioselective for Pro, Phe, and Trp.<sup>10</sup> Then, Pu and co-workers synthesized and investigated the binaphthol derivatives in the fluorescent recognition of amino acid derivatives.<sup>11</sup> In 2007, our group reported that compounds (*R,S*)-**1** and (*R,S*)-**2** acted as excellent agents for asymmetric catalysis.<sup>12</sup>

Herein, we describe that compound (*R,S*)-**1** shows high enantioselectivity in the fluorescent recognition of a series of amino acid derivatives. Our initial work began with the interaction of the sensor (*R,S*)-**1** (Scheme 1) and *N*-Boc-proline (**3**). Recognition **3** was carried out in solutions such as CH<sub>3</sub>CN, THF, and MeOH, and no

enantioselectivity could be found. Although CH<sub>2</sub>Cl<sub>2</sub> was used as the solvent, there was a slight discrimination between *D*-**3** and *L*-**3**.

Then, chiral recognition was investigated under the condition of CH<sub>2</sub>Cl<sub>2</sub>/hexane (3:7) containing 1.2% THF. THF was added to improve the solubility of **3**. And adding 20% THF had little influence on the whole chiral recognition system. The sensor (*R,S*)-**1** was



Scheme 1. Structure of chiral sensors.

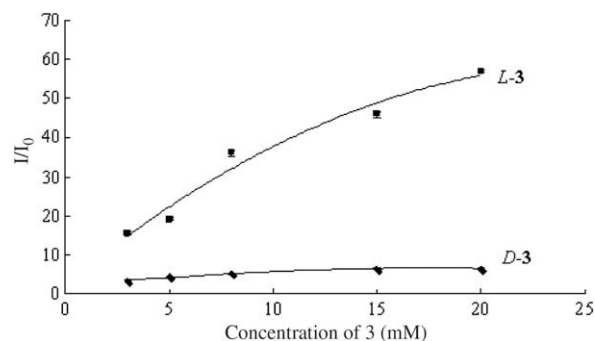
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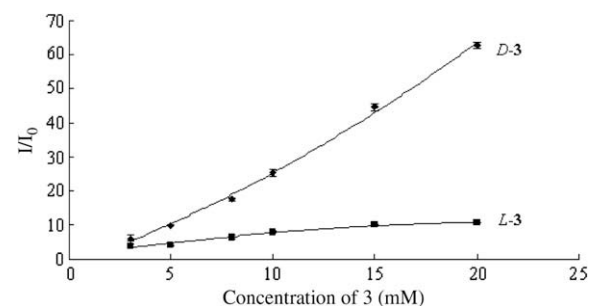
emitted at a short wavelength of 365 nm and a long wavelength of 434 nm on excitation at 278 nm. To our delight, it showed highly enantioselective fluorescent response to **3** (Fig. 1) at the short wavelength emission band. As shown in Figure 1, the enantiomer *L*-**3** ( $2 \times 10^{-2}$  M) enhanced the fluorescence intensity of (*R,S*)-**1** 10.4 times more than *D*-**3** ( $2 \times 10^{-2}$  M) did, that is,  $ef = 10.4^{11b}$  ( $ef$ : enantiomeric fluorescence difference ratio =  $(I_L - I_0)/(I_D - I_0)$ , where  $I_0$  is the fluorescence intensity of the receptor,  $I_D$  and  $I_L$  are the fluorescence intensities of the receptor in the presence of *D*-**3** or *L*-**3**). In addition, (*R,S*)-**2** and (*S,S*)-**2** were synthesized and used as sensors. Both *L*-**3** and *D*-**3** were found to enhance the fluorescence of (*R,S*)-**2** under optimized conditions. But the fluorescence enhancement did not indicate any enantioselectivity.

This result of the chiral recognition arose from three main factors: First, it was quite logical to infer from these experiments that the characteristic enantioselective fluorescence enhancement by the sensor molecule owed its origin to –NH group of sensors and –COOH group of *N*-Boc-amino acids. When –H was taken place by –CH<sub>3</sub>, it was more difficult for *N*-Boc-amino acids to complex well with the sensor (*R,S*)-**2** due to the steric hindrance effect. Second, high polar solution was not favorable for the hydrogen bond formation between sensor and *N*-Boc-amino acids. So, low polar solution was crucial for chiral recognition. Therefore, it was essential that hydrogen bonding interaction contributes for chiral recognition. Third, compound (*R,S*)-**1** contained nitrogen atoms adjacent to the naphthyl rings. And the lone pair of electrons of the nitrogen atom was available for photoinduced-electron-transfer (PET)<sup>13</sup> quenching of naphthyl rings. Similarly, the nitrogen atom in amino acids also quenches the fluorescence of the fluorophore. So we introduced the *N*-protecting group into the amino acids to suppress PET quenching.<sup>11b</sup> In the presence of *L*-**3** or *D*-**3**, (*R,S*)-**1** exhibited significant fluorescence enhancement owing to the suppressed PET quenching when the acidic proton of *N*-Boc-amino acid interacted with the nitrogen which (*R,S*)-**1** contained. Since the interaction of the sensor with the two enantiomers of *N*-Boc-proline would generate two diastereomers, different fluorescence enhancements occurred. Because there was the special cycle structure in *N*-Boc-proline which encouraged the interaction between sensors and *N*-Boc-proline, the result was excellent.

The effect of various concentrations of *D*-**3** or *L*-**3** on the fluorescence of (*R,S*)-**1** at 365 nm was investigated (Fig. 2). The error bars shown in Figure 2 were obtained by four independent measurements. These experiments demonstrated that the concentration ranged from  $3 \times 10^{-3}$  M to  $2 \times 10^{-2}$  M, and that the fluorescence



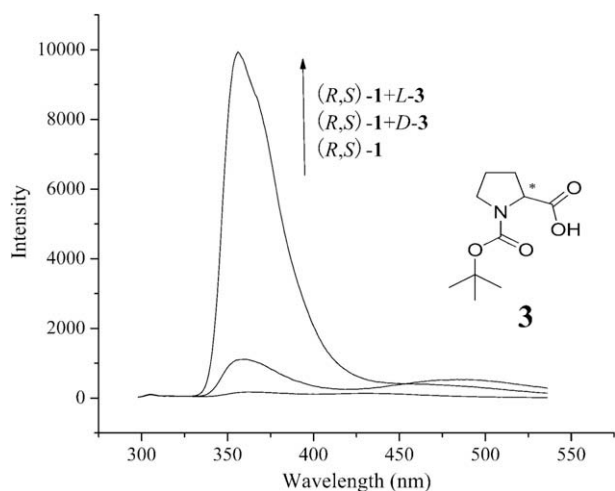
**Figure 2.** Fluorescence intensity change of (*R,S*)-**1** ( $1 \times 10^{-4}$  M in CH<sub>2</sub>Cl<sub>2</sub>/hexane (3:7), containing 1.2% THF) versus concentration of *D*-**3** and *L*-**3**.



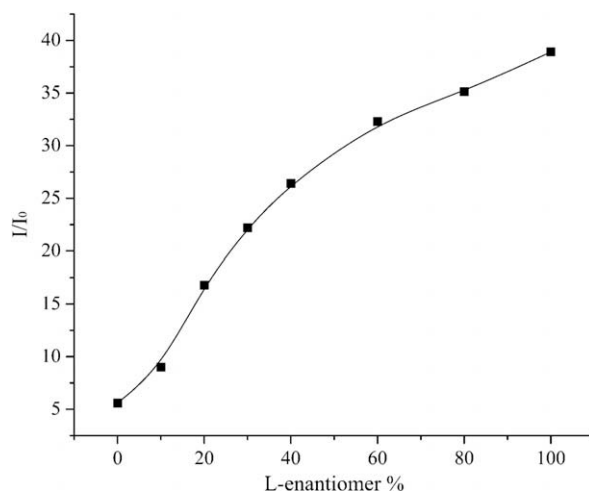
**Figure 3.** Fluorescence intensity change of (*S,S*)-**1** ( $1 \times 10^{-4}$  M in CH<sub>2</sub>Cl<sub>2</sub>/hexane (3:7), containing 1.2% THF) versus concentration of *D*-**3** and *L*-**3**.

intensity was enhanced. While *L*-**3** led to 16–57-fold increase in the fluorescence intensity of (*R,S*)-**1**, *D*-**3** caused little change in the fluorescence. Thus, the sensor (*R,S*)-**1** proved to be highly enantioselective toward **3**.

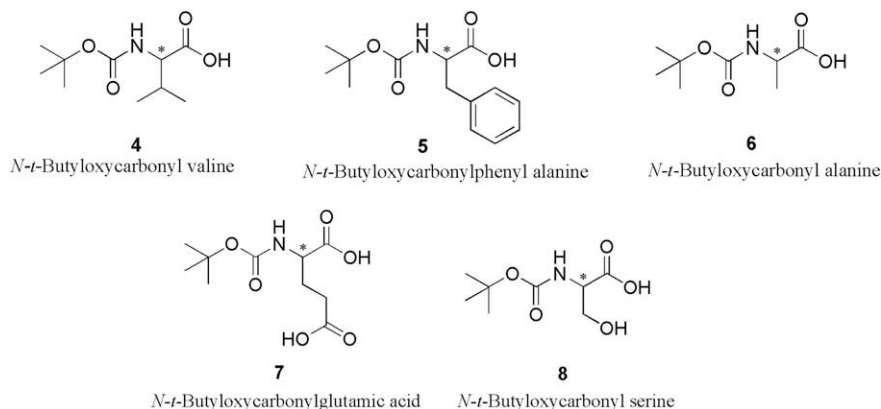
Then we changed the axial chirality of the naphthyl rings and studied the interaction of (*S,S*)-**1** with *D*-**3** or *L*-**3**. As expected, we noted a ‘reverse image’ relationship between the fluorescence enhancement of (*R,S*)-**1** and (*S,S*)-**1** in the presence of *N*-Boc-proline. And the  $ef$  observed for the interaction of (*S,S*)-**1** and **3** ( $ef = 6.6$ ) was not as high as that observed for the interaction of (*R,S*)-**1** and **3** ( $ef = 10.4$ ) under the same conditions. The error bars shown in Figure 3 were obtained by four independent measure-



**Figure 1.** Fluorescence spectra of (*R,S*)-**1** ( $1 \times 10^{-4}$  M in CH<sub>2</sub>Cl<sub>2</sub>/hexane (3:7), containing 1.2% THF) with/without *D*-**3** and *L*-**3** ( $2 \times 10^{-2}$  M) ( $\lambda_{exc} = 278$  nm, emission slits = 5.0 nm).



**Figure 4.** Fluorescence enhancement of (*R,S*)-**1** ( $1 \times 10^{-4}$  M in CH<sub>2</sub>Cl<sub>2</sub>/hexane (3:7), containing 1.2% THF) versus the enantiomeric composition of **3** ( $8 \times 10^{-3}$  M).



**Figure 5.** Structures of the amino acid derivatives 4–8.

**Table 1**

Results for the enantioselective fluorescent responses of (*R,S*)-**1** ( $5 \times 10^{-5}$  M) to the chiral amino acid derivatives **3–8**

Compound	Sensor	Solvent	Concentration (M)	Ef
<b>3</b>	( <i>R,S</i> )- <b>1</b>	CH <sub>2</sub> Cl <sub>2</sub> /hexane (3:7) containing 1.2% THF	$2 \times 10^{-2}$	10.4
<b>4</b>	( <i>R,S</i> )- <b>1</b>	CH <sub>2</sub> Cl <sub>2</sub> /hexane (3:7)	$5 \times 10^{-3}$	7.9
<b>5</b>	( <i>R,S</i> )- <b>1</b>	CH <sub>2</sub> Cl <sub>2</sub> /hexane (3:7)	$8 \times 10^{-3}$	4.0
<b>6</b>	( <i>R,S</i> )- <b>1</b>	CH <sub>2</sub> Cl <sub>2</sub> /hexane (3:7)	$8 \times 10^{-3}$	2.5
<b>7</b>	( <i>S,S</i> )- <b>1</b>	CH <sub>2</sub> Cl <sub>2</sub> /hexane (3:7) containing 4% THF	$8 \times 10^{-3}$	2.3
<b>8</b>	( <i>R,S</i> )- <b>1</b>	CH <sub>2</sub> Cl <sub>2</sub> /hexane (3:7) containing 4% THF	$8 \times 10^{-3}$	1.4

ments with the concentration ranging from  $3 \times 10^{-3}$  M to  $2 \times 10^{-2}$  M. This was confirmed by the enantioselective response of (*S,S*)-**1** and (*R,S*)-**1** towards the enantiomers of the **3**. And the axial chirality of naphthyl rings was considered to recognize chiral amino acid derivatives. The fluorescence intensity change of (*R,S*)-**1** with respect to the enantiomeric composition of **3** was investigated. As shown in Figure 4, the fluorescence intensity of (*R,S*)-**1** increased with the increasing *L*-component of **3**. Therefore, (*R,S*)-**1** might be used as a fluorescent sensor to readily determine the enantiomeric composition of **3**. Further investigation of the interaction of (*R,S*)-**1** or (*S,S*)-**1** with other *N*-Boc-amino acids **3–8** (Fig. 5) was performed. Table 1 summarizes the highest ef which was observed for the interaction of (*R,S*)-**1** or (*S,S*)-**1** with the two enantiomers of these *N*-Boc-amino acids. All these compounds enhanced the fluorescence of the binaphthyl fluorophore, respectively. Amino acid **7** showed the opposite enantioselective response to that of others.

In conclusion, we have developed a highly enantioselective fluorescent sensor for the chiral recognition of *N*-Boc-proline and other amino acid derivatives. It is observed that within a certain concentration range, *L*-*N*-Boc-proline could increase the fluorescence intensity of the sensor (*R,S*)-**1** by 16–57-fold, while *D*-*N*-Boc-proline led to only sixfold increase. It could be used in ascertaining the enantiomeric composition of *D*- and *L*-*N*-Boc-proline and in finding analytical applications over a definite concentration range.

**General procedure:** Materials: The *N*-Boc-proline and *N*-Boc-alanine were synthesized and recrystallized. Other *N*-Boc-amino acids were purchased from Aladdin. Compounds (*R,S*)-**1**, (*S,S*)-**1**, (*R,S*)-**2**, and (*S,S*)-**2** were synthesized according to the literature procedure.<sup>12</sup> All the solvents were redistilled. The general preparation procedure for the fluorescent measurement is described below:<sup>11b,14</sup> A mixture solvent stock solution of the chiral compound (*R,S*)-**1** was freshly prepared before each measurement. Different concentrations stock solution of the chiral *N*-Boc-amino acid in CH<sub>2</sub>Cl<sub>2</sub>/hexane (3:7), containing 1.2% THF, was freshly prepared. Then, the sensor solution was mixed with the *N*-Boc-amino acid

solution at room temperature in a 5 mL volumetric flask and diluted to the desired concentration. THF was added to improve the solubility of the *N*-Boc-amino acid. The resulting solution was allowed to stand at room temperature for 3 h and the fluorescence was measured on a Hitachi FL spectrophotometer (F-7000).

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